

Package ‘proteoQC’

April 10, 2015

Type Package

Title An R package for proteomics data quality control

Version 1.2.0

Author Bo Wen <wenbo@genomics.cn>, Laurent Gatto <lg390@cam.ac.uk>

Maintainer Bo Wen <wenbo@genomics.cn>

Description This package creates a HTML format QC report for MS/MS-based proteomics data. The report is intended to allow the user to quickly assess the quality of proteomics data.

Depends R (>= 3.0.0), XML, VennDiagram, MSnbase

Imports rTANDEM, plyr, seqinr, Nozzle.R1, ggplot2, reshape2, parallel,
Rcpp (>= 0.11.1)

LinkingTo Rcpp

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Suggests RforProteomics (>= 1.0.16), knitr, BiocStyle, rpx, R.utils,
RUnit, BiocGenerics

VignetteBuilder knitr

biocViews Proteomics, MassSpectrometry, QualityControl, Visualization,
ReportWriting

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addSummaryChart	<i>Add PRIDE summary charts</i>
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Description

Add PRIDE summary charts in the technical replicate level

Usage

```
addSummaryChart(res)
```

Arguments

res	An object returned by msQCpipe function
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calcMSQCMetrics	<i>Calculate the MS1 and MS2 level QC metrics</i>
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Description

Calculate the MS1 level QC metrics

Usage

```
calcMSQCMetrics(spectraList = NULL, cpu = 2, outdir = "./")
```

Arguments

- | | |
|-------------|---------------------------------|
| spectraList | An experiment design input file |
| cpu | The number of cpu used |
| outdir | Output directory |

Value

A data frame

Author(s)

Bo Wen <wenbo@genomics.cn>

chargeStat

Charge distribution

Description

Read the charge information from mgf file

Usage

```
chargeStat(mgf = NULL)
```

Arguments

- | | |
|-----|----------------|
| mgf | A file of mgf. |
|-----|----------------|

Value

A vector object

Author(s)

Bo Wen <wenbo@genomics.cn>

Examples

```
mgf.zip <- system.file("extdata/mgf.zip", package = "proteoQC")
unzip(mgf.zip)
charge <- chargeStat("test.mgf")
```

cntStat	<i>contaminants stat</i>
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Description

Common Contaminants in Proteomics Mass Spectrometry Experiments

Usage

```
cntStat(res)
```

Arguments

res	An object of msQCres
-----	----------------------

Value

A data.frame will be shown in HTML report

combineRun	<i>Combine multiple results</i>
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Description

Combine multiple results

Usage

```
combineRun(pepFiles, fasta, outPathFile, outdir, prefix)
```

Arguments

pepFiles	peptideSummary files
fasta	database file
outPathFile	out file
outdir	output directory
prefix	output prefix

Value

A data.frame

Author(s)

Bo Wen <wenbo@genomics.cn>

`createTargetDecoyDB` *Create target-decoy database*

Description

Create target-decoy database

Usage

`createTargetDecoyDB(fa, outdb)`

Arguments

<code>fa</code>	target database
<code>outdb</code>	output target-decoy database

Value

target-decoy database file name

Author(s)

Bo Wen <wenbo@genomics.cn>

`getEnzyme` *Get the enzymes list*

Description

Get the enzymes list

Usage

`getEnzyme()`

Value

A data frame which contains all of the enzymes

Author(s)

Bo Wen <wenbo@genomics.cn>

`getMods` *Get the modification list*

Description

Get the modification list

Usage

```
getMods()
```

Value

A data frame which contains all of the modifications

Author(s)

Bo Wen <wenbo@genomics.cn>

`labelRatio` *Calculate the labeling efficiency of isobaric labeling data*

Description

Calculate the labeling efficiency of isobaric labeling data

Usage

```
labelRatio(ms = NULL, iClass = 1, delta = 0.05)
```

Arguments

<code>ms</code>	MS/MS file.
<code>iClass</code>	Isobaric tag class, 1=iTRAQ-8plex.
<code>delta</code>	The mass error for reporter matching.

Value

A vector object

Author(s)

Bo Wen <wenbo@genomics.cn>

Examples

```
mgf.zip <- system.file("extdata/mgf.zip", package = "proteoQC")
unzip(mgf.zip)
a <- labelRatio("test.mgf")
```

loadmsQCres

*Load the result of msQCpipe***Description**

Load the result of [msQCpipe](#)

Usage

```
loadmsQCres(outdir)
```

Arguments

outdir	The output directory of msQCpipe
--------	--

Author(s)

Laurent Gatto <lg390@cam.ac.uk>, Bo Wen <wenbo@genomics.cn>

Examples

```
zqc <- system.file("extdata/qc.zip", package = "proteoQC")
unzip(zqc)
qcres <- loadmsQCres("./qc")
```

msQCpipe

*The main function of msQC pipeline***Description**

This function is designed to automate generating of target-decoy database, database searching and post-processing.

Usage

```
msQCpipe(spectralist = NULL, fasta = "", outdir = "./", mode = "",
miss = 2, enzyme = 1, varmod = NULL, fixmod = NULL, tol = 10,
tolu = "ppm", itol = 0.6, itolu = "Daltons", threshold = 0.01,
cpu = 0, xmx = 2, ...)
```

Arguments

<code>spectralist</code>	A file contains the experiment design
<code>fasta</code>	database file, must contain decoy sequences
<code>outdir</code>	output directory
<code>mode</code>	identification or quantification
<code>miss</code>	max miss cleavage
<code>enzyme</code>	enzyme
<code>varmod</code>	Variable modifications are those which may or may not be present.
<code>fixmod</code>	Fixed modifications are applied universally, to every instance of the specified residue(s) or terminus.
<code>tol</code>	The error window on experimental peptide mass values
<code>tolu</code>	Units can be selected from: ppm, Daltons(also da or Da).
<code>itol</code>	Error window for MS/MS fragment ion mass values.
<code>itolu</code>	Units can be selected from: Daltons(also da or Da)
<code>threshold</code>	FDR value for PSM
<code>cpu</code>	Max number of cpu used
<code>xmx</code>	JAVA -Xmx
<code>...</code>	Additional parameters passed to <code>read.table</code> used to read the experimental design.

Value

A list which contains all of the information for data quality report generating

Author(s)

Bo Wen <wenbo@genomics.cn>

Examples

```
## Not run:
library("rpx")
px <- PXDataset("PXD000864")
mgfs <- grep("mgf", pxfilenames(px), value = TRUE)
mgfs <- grep("-0[5-6]-[1|2]", mgfs, value=TRUE)
mgfffiles <- pxget(px, mgfs)
library("R.utils")
mgfffiles <- sapply(mgfffiles, gunzip)
## Generate the lightweight qc report,
## trim the mgf files to 1/10 of their size.
trimMgf <- function(f, m = 1/10, overwrite = FALSE) {
  message("Reading ", f)
  x <- readLines(f)
  beg <- grep("BEGIN IONS", x)
  end <- grep("END IONS", x)
```

```

n <- length(beg)
message("Sub-setting to ", m)
i <- sort(sample(n, floor(n * m)))
k <- unlist(mapply(seq, from = beg[i], to = end[i]))
if (overwrite) {
  unlink(f)
  message("Writing ", f)
  writeLines(x[k], con = f)
  return(f)
} else {
  g <- sub(".mgf", "_small.mgf", f)
  message("Writing ", g)
  writeLines(x[k], con = g)
  return(g)
}
set.seed(1)
mgfffiles <- sapply(mgfffiles, trimMgf, overwrite = TRUE)
fas <- pxget(px, "TTE2010.zip")
fas <- unzip(fas)
design <- system.file("extdata/PXD000864-design.txt", package = "proteoQC")
read.table(design, header = TRUE)
qcres <- msQCpipe(spectralist = design,
                    fasta = fas,
                    outdir = "./qc",
                    miss = 0,
                    enzyme = 1, varmod = 2, fixmod = 1,
                    tol = 10, itol = 0.6, cpu = 2,
                    mode = "identification")
html <- reportHTML(qcres)

## End(Not run)

```

plotBioRepVenn

*Venn plot in biological replicate level***Description**

Venn plot in biological replicate level

Usage

```
plotBioRepVenn(res)
```

Arguments

res	An object of msQCres
-----	----------------------

Value

The name of the figure

plotFractionIDResult *Barplot in different level for each fraction*

Description

Barplot in different level for each fraction

Usage

```
plotFractionIDResult(res, level = NA)
```

Arguments

res	An object of msQCres
level	1: total spectrum, 2: identified spectrum, 3: identified peptide, 4: identified protein.

Value

The name of the figure

plotMS1Error *plot MS1 mass error*

Description

plot MS1 mass error

Usage

```
plotMS1Error(res, plot.class = "ppm")
```

Arguments

res	An object of msQCres
plot.class	ppm or da

Value

The name of the figure

`plotMS2Error` *plot MS2 mass error*

Description

plot MS2 mass error

Usage

`plotMS2Error(res)`

Arguments

`res` An object of `msQCres`

Value

The name of the figure

`plotMS2Error_obsolete` *plot MS2 mass error*

Description

plot MS2 mass error

Usage

`plotMS2Error_obsolete(res)`

Arguments

`res` An object of `msQCres`

Value

The name of the figure

```
plotSampleIDResultErrorBar
```

Error barplot in different level for each fraction

Description

Error Barplot in different level for each fraction

Usage

```
plotSampleIDResultErrorBar(res, level = NA)
```

Arguments

res	An object of parser result
level	1: total spectrum, 2: identified spectrum, 3: identified peptide, 4: identified protein.

Value

The name of the figure

```
plotSampleVenn
```

Venn plot in sample level

Description

Venn plot in sample level

Usage

```
plotSampleVenn(res)
```

Arguments

res	An object of msQCres
-----	----------------------

Value

The name of the figure

plotTechRepVenn *Venn plot in technical replicate level*

Description

Venn plot in technical replicate level

Usage

```
plotTechRepVenn(res)
```

Arguments

res An object of msQCres

Value

The name of the figure

print.msQCres *Print the information of msQCres object*

Description

Print the information of msQCres object

Usage

```
## S3 method for class msQCres  
print(x, ...)
```

Arguments

x A msQCres object
... Additional parameters

Author(s)

Laurent Gatto <lg390@cam.ac.uk>, Bo Wen <wenbo@genomics.cn>

Examples

```
zpqc <- system.file("extdata/qc.zip", package = "proteoQC")  
unzip(zpqc)  
qcres <- loadmsQCres("./qc")  
print.msQCres(qcres)
```

proteinGroup*Protein inference***Description**

Protein inference

Usage

```
proteinGroup(file = NULL, db = "", pepColName = "peptide",
            proColName = "protein", spectrumColName = "index", proSep = ";",
            outfile = NULL, xmx = 1)
```

Arguments

<code>file</code>	A file containing the information of peptides to proteins.
<code>db</code>	A protein database of fasta format.
<code>pepColName</code>	The column name of peptide sequence.
<code>proColName</code>	The column name of protein ID.
<code>spectrumColName</code>	The column name of spectrum index.
<code>proSep</code>	The separator of protein ID, default is "".
<code>outfile</code>	The output file name of protein group result.
<code>xmx</code>	JAVA -Xmx, default is 1.

Author(s)

Bo Wen <wenbo@genomics.cn>

Examples

```
pep.zip <- system.file("extdata/pep.zip", package = "proteoQC")
unzip(pep.zip)
proteinGroup(file = "pep.txt", outfile = "pg.txt")
```

reportHTML	<i>HTML format report generator</i>
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Description

HTML format report generator

Usage

```
reportHTML(res)
```

Arguments

res	An object returned by msQCpipe function
-----	---

Value

null

Author(s)

Bo Wen <wenbo@genomics.cn>

Examples

```
zpzq <- system.file("extdata/qc.zip", package = "proteoQC")
unzip(zpzq)
qcres <- loadmsQCres("./qc")
html <- reportHTML(qcres)
```

runTandem	<i>Run X!Tandem</i>
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Description

Run X!Tandem

Usage

```
runTandem(spectra = "", fasta = "", outdir = "./", outprefix = "",
cpu = 1, enzyme = 1, xmx = 2, varmod = NULL, fixmod = NULL,
tol = 10, tolu = "ppm", itol = 0.6, itolu = "Daltons", miss = 1)
```

Arguments

spectra	MS/MS peak list file
fasta	database file
outdir	output directory
outprefix	output file prefix
cpu	The number of CPU used for X!Tandem
enzyme	The ID of enzyme used for database searching. See showEnzyme .
xmx	Set for parameter of "Java -Xmx".
varmod	Variable modifications used for database searching. See showMods .
fixmod	Fixed modifications used for database searching. See showMods .
tol	The error window on experimental peptide mass values
tolu	Units can be selected from: ppm, Daltons.
itol	Error window for MS/MS fragment ion mass values.
itolu	Units can be selected from: Daltons
miss	Max miss cleavage

Value

a file path

Author(s)

Bo Wen <wenbo@genomics.cn>

`showEnzyme`

Show all enzymes

Description

Shown all enzymes

Usage

`showEnzyme()`

Value

A data frame which contains all of the enzymes

Author(s)

Bo Wen <wenbo@genomics.cn>

Examples

`showEnzyme()`

<code>showMods</code>	<i>Shown all modifications</i>
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Description

Shown all modifications

Usage

`showMods()`

Value

A data frame which contains all of the modifications

Author(s)

Bo Wen <wenbo@genomics.cn>

Examples

`showMods()`

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