# Package 'MethylMix'

July 17, 2025

Title MethylMix: Identifying methylation driven cancer genes

Version 2.39.0

Description MethylMix is an algorithm implemented to identify hyper and hypomethylated genes for a disease. MethylMix is based on a beta mixture model to identify methylation states and compares them with the normal DNA methylation state. MethylMix uses a novel statistic, the Differential Methylation value or DM-value defined as the difference of a methylation state with the normal methylation state. Finally, matched gene expression data is used to identify, besides differential, functional methylation states by focusing on methylation changes that effect gene expression. References:

Gevaert 0. MethylMix: an R package for identifying DNA methylation-driven genes. Bioinformatics (Oxford, England). 2015;31(11):1839-41. doi:10.1093/bioinformatics/btv020.

Gevaert O, Tibshirani R, Plevritis SK. Pancancer analysis of DNA methylation-driven genes using MethylMix. Genome Biology. 2015;16(1):17. doi:10.1186/s13059-014-0579-8

**Depends** R (>= 3.2.0)

License GPL-2

**Encoding** UTF-8

LazyData true

Author Olivier Gevaert

Maintainer Olivier Gevaert <olivier.gevaert@gmail.com>

Type Package

**Date** 2018-07-13

**Imports** foreach, RPMM, RColorBrewer, ggplot2, RCurl, impute, data.table, limma, R.matlab, digest

Suggests BiocStyle, doParallel, testthat, knitr, rmarkdown

biocViews

DNAMethylation, Statistical Method, Differential Methylation, Gene Regulation, Gene Expression, Methylation Array, Differential Methylation, Gene Expression, Gene E

RoxygenNote 6.0.1

VignetteBuilder knitr

git\_url https://git.bioconductor.org/packages/MethylMix

git\_branch devel

2 Contents

git_last_commit f4d5f08
git_last_commit_date 2025-04-15
Repository Bioconductor 3.22
<b>Date/Publication</b> 2025-07-16

# **Contents**

Index

BatchData
betaEst_2
blc_2 3
ClusterProbes
ComBat_NoFiles
combineForEachOutput
Download_DNAmethylation
Download_GeneExpression
GEcancer
GetData
get_firehoseData
METcancer
MethylMix
MethylMix_MixtureModel
MethylMix_ModelGeneExpression
MethylMix_ModelSingleGene
MethylMix_PlotModel
MethylMix_Predict
MethylMix_RemoveFlipOver
METnormal
predictOneGene
Preprocess_CancerSite_Methylation27k
Preprocess_CancerSite_Methylation450k
Preprocess_DNAmethylation
Preprocess_GeneExpression
Preprocess_MAdata_Cancer
Preprocess_MAdata_Normal
ProbeAnnotation
SNPprobes
TCGA_BatchCorrection_MolecularData
TCGA_GENERIC_BatchCorrection
TCGA_GENERIC_CheckBatchEffect
TCGA_GENERIC_CleanUpSampleNames
TCGA_GENERIC_GetSampleGroups
TCGA_GENERIC_LoadIlluminaMethylationData
TCGA_GENERIC_MergeData
TCGA_GENERIC_MET_ClusterProbes_Helper_ClusterGenes_with_hclust 28
TCGA_Load_MolecularData
TCGA_Process_EstimateMissingValues

**30** 

BatchData 3

# Description

Data set with batch number for TCGA samples.

```
betaEst_2 The betaEst_2 function
```

# Description

Internal. Estimates a beta distribution via Maximum Likelihood. Adapted from RPMM package.

### Usage

```
betaEst_2(Y, w, weights)
```

# **Arguments**

Y data vector.

w posterior weights.weights Case weights.

### Value

(a,b) parameters.

```
blc_2 The blc_2 function
```

# Description

Internal. Fits a beta mixture model for any number of classes. Adapted from RPMM package.

### Usage

```
blc_2(Y, w, maxiter = 25, tol = 1e-06, weights = NULL, verbose = TRUE)
```

# **Arguments**

Y Data matrix (n x j) on which to perform clustering.
w Initial weight matrix (n x k) representing classification.

maxiter Maximum number of EM iterations.

tol Convergence tolerance.

weights Case weights.
verbose Verbose output.

4 ClusterProbes

#### Value

A list of parameters representing mixture model fit, including posterior weights and log-likelihood.

ClusterProbes The ClusterProbes function

# **Description**

This function uses the annotation for Illumina methylation arrays to map each probe to a gene. Then, for each gene, it clusters all its CpG sites using hierchical clustering and Pearson correlation as distance and complete linkage. If data for normal samples is provided, only overlapping probes between cancer and normal samples are used. Probes with SNPs are removed. This function is prepared to run in parallel if the user registers a parallel structure, otherwise it runs sequentially. This function also cleans up the sample names, converting them to the 12 digit format.

### Usage

```
ClusterProbes(MET_Cancer, MET_Normal, CorThreshold = 0.4)
```

### **Arguments**

MET\_Cancer data matrix for cancer samples.

MET\_Normal data matrix for normal samples.

CorThreshold correlation threshold for cutting the clusters.

# Value

List with the clustered data sets and the mapping between probes and genes.

# **Examples**

```
## Not run:
# Optional register cluster to run in parallel
library(doParallel)
cl <- makeCluster(5)
registerDoParallel(cl)

# Methylation data for ovarian cancer
cancerSite <- "OV"
targetDirectory <- paste0(getwd(), "/")

# Downloading methylation data
METdirectories <- Download_DNAmethylation(cancerSite, targetDirectory, TRUE)

# Processing methylation data
METProcessedData <- Preprocess_DNAmethylation(cancerSite, METdirectories)

# Saving methylation processed data
saveRDS(METProcessedData, file = paste0(targetDirectory, "MET_", cancerSite, "_Processed.rds"))
# Clustering methylation data</pre>
```

ComBat\_NoFiles 5

```
res <- ClusterProbes(METProcessedData[[1]], METProcessedData[[2]])
# Saving methylation clustered data
toSave <- list(METcancer = res[[1]], METnormal = res[[2]], ProbeMapping = res$ProbeMapping)
saveRDS(toSave, file = paste0(targetDirectory, "MET_", cancerSite, "_Clustered.rds"))
stopCluster(cl)
## End(Not run)</pre>
```

ComBat\_NoFiles

The ComBat\_NoFiles function

# Description

Internal. Performs batch correction.

# Usage

```
ComBat_NoFiles(dat, saminfo, type = "txt", write = F, covariates = "all",
    par.prior = F, filter = F, skip = 0, prior.plots = T)
```

# **Arguments**

dat	dat
saminfo	saminfo
type	currently supports two data file types 'txt' for a tab-delimited text file and 'csv' for an Excel .csv file (sometimes R handles the .csv file better, so use this if you have problems with a .txt file!).
write	if 'T' ComBat writes adjusted data to a file, and if 'F' and ComBat outputs the adjusted data matrix if 'F' (so assign it to an object! i.e. NewData <- ComBat('my expression.xls','Sample info file.txt', write=F)).
covariates	'covariates=all' will use all of the columns in your sample info file in the modeling (except array/sample name), if you only want use a some of the columns in your sample info file, specify these columns here as a vector (you must include the Batch column in this list).
par.prior	if 'T' uses the parametric adjustments, if 'F' uses the nonparametric adjustments—if you are unsure what to use, try the parametric adjustments (they run faster) and check the plots to see if these priors are reasonable.
filter	'filter=value' filters the genes with absent calls in > 1-value of the samples. The defaut here (as well as in dchip) is .8. Filter if you can as the EB adjustments work better after filtering. Filter must be numeric if your expression index file contains presence/absence calls (but you can set it >1 if you don't want to filter any genes) and must be 'F' if your data doesn't have presence/absence calls;
skip	is the number of columns that contain probe names and gene information, so 'skip=5' implies the first expression values are in column 6
prior.plots	if true will give prior plots with black as a kernal estimate of the empirical batch effect density and red as the parametric estimate.

#### Value

Results.

combineForEachOutput The combineForEachOutput function

#### **Description**

Internal. Function to combine results from the foreach loop.

# Usage

```
combineForEachOutput(out1, out2)
```

### **Arguments**

out1 result from one foreach loop.
out2 result from another foreach loop.

#### Value

List with the combined results.

Download\_DNAmethylation

 $The\ Download\_DNA methylation\ function$ 

# **Description**

Downloads DNA methylation data from TCGA.

### Usage

Download\_DNAmethylation(CancerSite, TargetDirectory, downloadData = TRUE)

# Arguments

CancerSite character of length 1 with TCGA cancer code.

TargetDirectory

character with directory where a folder for downloaded files will be created.

downloadData logical indicating if data should be downloaded (default: TRUE). If false, the

url of the desired data is returned.

# Value

list with paths to downloaded files for both 27k and 450k methylation data.

#### **Examples**

```
## Not run:
# Optional register cluster to run in parallel
library(doParallel)
cl <- makeCluster(5)</pre>
registerDoParallel(cl)
# Methylation data for ovarian cancer
cancerSite <- "OV"
targetDirectory <- paste0(getwd(), "/")</pre>
# Downloading methylation data
METdirectories <- Download_DNAmethylation(cancerSite, targetDirectory, TRUE)</pre>
# Processing methylation data
METProcessedData <- Preprocess_DNAmethylation(cancerSite, METdirectories)</pre>
# Saving methylation processed data
saveRDS(METProcessedData, file = paste0(targetDirectory, "MET_", cancerSite, "_Processed.rds"))
# Clustering methylation data
res <- ClusterProbes(METProcessedData[[1]], METProcessedData[[2]])</pre>
# Saving methylation clustered data
toSave <- list(METcancer = res[[1]], METnormal = res[[2]], ProbeMapping = res$ProbeMapping)
saveRDS(toSave, file = paste0(targetDirectory, "MET_", cancerSite, "_Clustered.rds"))
stopCluster(cl)
## End(Not run)
```

Download\_GeneExpression

The Download\_GeneExpression function

### **Description**

Downloads gene expression data from TCGA.

#### Usage

```
Download_GeneExpression(CancerSite, TargetDirectory, downloadData = TRUE)
```

### **Arguments**

CancerSite character of length 1 with TCGA cancer code.

TargetDirectory

character with directory where a folder for downloaded files will be created.

downloadData logical indicating if data should be downloaded (default: TRUE). If false, the url of the desired data is returned.

8 GEcancer

#### **Details**

This function downloads RNAseq data (file tag "mRNAseq\_Preprocess.Level\_3"), with the exception for OV and GBM, for which micro array data is downloaded since there is not enough RNAseq data

#### Value

list with paths to downloaded files for both 27k and 450k methylation data.

# **Examples**

```
## Not run:
 # Optional register cluster to run in parallel
 library(doParallel)
 cl <- makeCluster(5)</pre>
 registerDoParallel(cl)
 # Gene expression data for ovarian cancer
 cancerSite <- "OV"
 targetDirectory <- paste0(getwd(), "/")</pre>
 # Downloading gene expression data
 GEdirectories <- Download_GeneExpression(cancerSite, targetDirectory, TRUE)
 # Processing gene expression data
 GEProcessedData <- Preprocess_GeneExpression(cancerSite, GEdirectories)</pre>
 # Saving gene expression processed data
 saveRDS(GEProcessedData, file = paste0(targetDirectory, "GE_", cancerSite, "_Processed.rds"))
 stopCluster(cl)
 ## End(Not run)
GEcancer
                          Cancer Gene expression data of glioblastoma patients from the TCGA
                          project
```

#### **Description**

Cancer Gene expression data of glioblastoma patients from the TCGA project. A set of 14 genes that have been shown in the literature to be involved in differential methylation in glioblastoma were selected as an example to try out MethylMix.

### Usage

```
data(GEcancer)
```

#### **Format**

A numeric matrix with 14 rows (genes) and 251 columns (samples).

GetData 9

#### References

Cancer Genome Atlas Research Network. Comprehensive genomic characterization defines human glioblastoma genes and core pathways. Nature. 2008 Oct 23; 455(7216):1061-8. doi: 10.1038/nature07385. Epub 2008 Sep 4. Erratum in: Nature. 2013 Feb 28;494(7438):506. PubMed PMID: 18772890; PubMed Central PMCID: PMC2671642.

#### See Also

TCGA: The Cancer Genome Atlas: http://cancergenome.nih.gov/

GetData

The GetData function

# Description

This function wraps the functions for downloading and pre-processing DNA methylation and gene expression data, as well as for clustering CpG probes.

### Usage

GetData(cancerSite, targetDirectory)

### **Arguments**

cancerSite character of length 1 with TCGA cancer code. targetDirectory

character with directory where a folder for downloaded files will be created.

#### **Details**

Pre-process of DNA methylation data includes eliminating samples and genes with too many NAs, imputing NAs, and doing Batch correction. If there is both 27k and 450k data, and both data sets have more than 50 samples, we combine the data sets, by reducing the 450k data to the probes present in the 27k data, and bath correction is performed again to the combined data set. If there are samples with both 27k and 450k data, the 450k data is used and the 27k data is discarded, before the step mentioned above. If the 27k or the 450k data does not have more than 50 samples, we use the one with the greatest number of samples, we do not combine the data sets.

For gene expression, this function downloads RNAseq data (file tag "mRNAseq\_Preprocess.Level\_3"), with the exception for OV and GBM, for which micro array data is downloaded since there is not enough RNAseq data. Pre-process of gene expression data includes eliminating samples and genes with too many NAs, imputing NAs, and doing Batch correction.

For the clustering of the CpG probes, this function uses the annotation for Illumina methylation arrays to map each probe to a gene. Then, for each gene, it clusters all its CpG sites using hierchical clustering and Pearson correlation as distance and complete linkage. If data for normal samples is provided, only overlapping probes between cancer and normal samples are used. Probes with SNPs are removed.

This function is prepared to run in parallel if the user registers a parallel structure, otherwise it runs sequentially.

This function also cleans up the sample names, converting them to the 12 digit format.

10 get\_firehoseData

#### Value

The following files will be created in target directory:

- gdac: a folder with the raw data downloaded from TCGA.
- MET\_CancerSite\_Processed.rds: processed methylation data at the CpG sites level (not clustered).
- GE\_CancerSite\_Processed.rds: processed gene expression data.
- data\_CancerSite.rds: list with both gene expression and methylation data. Methylation data is clustered and presented at the gene level. A matrix with the mapping from CpG sites to genes is included.

# **Examples**

```
## Not run:
# Get data for ovarian cancer
cancerSite <- "OV"
targetDirectory <- paste0(getwd(), "/")
GetData(cancerSite, targetDirectory)

# Optional register cluster to run in parallel
library(doParallel)
cl <- makeCluster(5)
registerDoParallel(cl)

cancerSite <- "OV"
targetDirectory <- paste0(getwd(), "/")
GetData(cancerSite, targetDirectory)

stopCluster(cl)

## End(Not run)</pre>
```

get\_firehoseData

The get\_firehoseData function

#### **Description**

Gets data from TCGA's firehose.

### Usage

```
get_firehoseData(downloadData = TRUE, saveDir = "./",
   TCGA_acronym_uppercase = "LUAD", dataType = "stddata",
   dataFileTag = "mRNAseq_Preprocess.Level_3", FFPE = FALSE,
   fileType = "tar.gz", gdacURL = "http://gdac.broadinstitute.org/runs/",
   untarUngzip = TRUE, printDisease_abbr = FALSE)
```

METcancer 11

#### **Arguments**

downloadData logical indicating if data should be downloaded (default: TRUE). If false, the

url of the desired data is returned.

saveDir path to directory to save downloaded files.

TCGA\_acronym\_uppercase

TCGA's cancer site code.

dataType type of data in TCGA (default: "stddata").

dataFileTag name of the file to be downloaded (the default is to download RNAseq data, but

this can be changed to download other data).

FFPE logical indicating if FFPE data should be downloaded (default: FALSE).

fileType type of downloaded file (default: "fileType", other type not admitted at the mo-

ment).

gdacURL gdac url.

untarUngzip logical indicating if the gzip file downloaded should be untarred (default: TRUE).

printDisease\_abbr

if TRUE data is not downloaded but all the possible cancer sites codes are shown

(default: FALSE).

#### Value

DownloadedFile path to directory with downloaded files.

METcancer DNA methylation data from cancer tissue from glioblastoma patients from the TCGA project

#### **Description**

Cancer Gene expression data of glioblastoma patients from the TCGA project. A set of 14 genes that have been shown in the literature to be involved in differential methylation in glioblastoma were selected as an example to try out MethylMix.

### Usage

data(METcancer)

#### **Format**

A numeric matrix with 14 rows (genes) and 251 columns (samples).

### References

Cancer Genome Atlas Research Network. Comprehensive genomic characterization defines human glioblastoma genes and core pathways. Nature. 2008 Oct 23; 455(7216):1061-8. doi: 10.1038/nature07385. Epub 2008 Sep 4. Erratum in: Nature. 2013 Feb 28;494(7438):506. PubMed PMID: 18772890; PubMed Central PMCID: PMC2671642.

#### See Also

TCGA: The Cancer Genome Atlas: http://cancergenome.nih.gov/

12 MethylMix

MethylMix	MethylMix: Mixture model for DNA methylation data in cancer.

### **Description**

MethylMix identifies DNA methylation driven genes by modeling DNA methylation data in cancer vs. normal and looking for homogeneous subpopulations. In addition matched gene expression data (e.g. from microarray technology or RNA sequencing) is used to identify functional DNA methylation events by requiring a negative correlation between methylation and gene expression of a particular gene. See references below.

# Usage

```
MethylMix(METcancer, GEcancer, METnormal = NULL, listOfGenes = NULL,
filter = TRUE, NoNormalMode = FALSE, OutputRoot = "")
```

### **Arguments**

METcancer	Matrix with the methylation data of cancer tissue with genes in rows and samples in columns.
GEcancer	Gene expression data for cancer tissue with genes in rows and samples in columns.
METnormal	Matrix with the normal methylation data of the same genes as in METcancer. Again genes in rows and samples in columns. The samples do not have to match with the cancer data. If this argument is NULL, MethylMix will run without comparing to normal samples.
listOfGenes	Vector with genes names to be evaluated, names must coincide with the names of the rows of METcancer.
filter	Logical indicating if the linear regression to select genes with significative linear negative relation between methylation and gene expression should be performed (default: TRUE).
NoNormalMode	Logical indicating if the methylation states found in the cancer samples should be compared to the normal samples (default: FALSE).
OutputRoot	Path to store the MethylMix results object.

### Value

MethylMixResults is a list with the following components:

MethylationDrivers

Genes identified as transcriptionally predictive and differentially methylated by MethylMix.

NrComponents The number of methylation states found for each driver gene.

MixtureStates A list with the DM-values for each driver gene. Differential Methylation values

(DM-values) are defined as the difference between the methylation mean in one mixture component of cancer samples and the methylation mean in the normal

samples, for a given gene.

MethylationStates

Matrix with DM-values for all driver genes (rows) and all samples (columns).

Classifications

Matrix with integers indicating to which mixture component each cancer sample was assigned to, for each gene.

Models

Beta mixture model parameters for each driver gene.

#### References

Gevaert 0. MethylMix: an R package for identifying DNA methylation-driven genes. Bioinformatics (Oxford, England). 2015;31(11):1839-41. doi:10.1093/bioinformatics/btv020.

Gevaert O, Tibshirani R, Plevritis SK. Pancancer analysis of DNA methylation-driven genes using MethylMix. Genome Biology. 2015;16(1):17. doi:10.1186/s13059-014-0579-8.

Pierre-Louis Cedoz, Marcos Prunello, Kevin Brennan, Olivier Gevaert; MethylMix 2.0: an R package for identifying DNA methylation genes. Bioinformatics. doi:10.1093/bioinformatics/bty156.

### **Examples**

```
# load the three data sets needed for MethylMix
data(METcancer)
data(METnormal)
data(GEcancer)

# run MethylMix on a small set of example data
MethylMixResults <- MethylMix(METcancer, GEcancer, METnormal)

## Not run:
# run in parallel
library(doParallel)
cl <- makeCluster(5)
registerDoParallel(cl)
MethylMixResults <- MethylMix(METcancer, GEcancer, METnormal)
stopCluster(cl)

## End(Not run)</pre>
```

MethylMix\_MixtureModel

The MethylMix\_MixtureModel function

# Description

Internal. Prepares all the structures to store the results and calls in a foreach loop a function that fits the mixture model in each gene.

# Usage

```
MethylMix_MixtureModel(METcancer, METnormal = NULL, FunctionalGenes,
   NoNormalMode = FALSE)
```

#### **Arguments**

METcancer matrix with methylation data for cancer samples (genes in rows, samples in

columns).

METnormal matrix with methylation data for normal samples (genes in rows, samples in

columns). If NULL no comparison to normal samples will be done.

FunctionalGenes

vector with genes names to be considered for the mixture models.

NoNormalMode logical, if TRUE no comparison to normal samples is performed. Defaults to

FALSE.

#### Value

MethylationStates matrix of DM values, with driver genes in the rows and samples in the columns.

NrComponents matrix with the number of components identified for each driver gene.

Models list with the mixture model fitted for each driver gene.

MethylationDrivers character vector with the genes found by MethylMix as differentially methylated and transcriptionally predictive (driver genes).

MixtureStates a list with a matrix for each driver gene containing the DM values.

Classifications a vector indicating to which component each sample was assigned.

MethylMix\_ModelGeneExpression

The MethylMix\_ModelGeneExpression function

# **Description**

Model gene expression as a function of gene expression with a simple linear regression model. Genes with a significant negative linear association between DNA methylation and gene expression are returned.

# Usage

MethylMix\_ModelGeneExpression(METcancer, GEcancer, CovariateData = NULL)

# Arguments

METcancer matrix with methylation data for cancer samples (genes in rows, samples in

columns).

GEcancer matrix with gene expression data for cancer samples (genes in rows, samples in

columns).

CovariateData vector (numeric or character) indicating a covariate to be included in the model

to adjust for it. Not used in an standard run of MethylMix. It can be used if

samples can from different tissue type, for example.

#### Value

vector with the names of the genes for which there is a significant linear and negative association between methylation and gene expression.

#### **Examples**

```
# load data sets
data(METcancer)
data(GEcancer)

# model gene expression
MethylMixResults <- MethylMix_ModelGeneExpression(METcancer, GEcancer)</pre>
```

MethylMix\_ModelSingleGene

The MethylMix\_ModelSingleGene function

## **Description**

Internal. For a given gene, this function fits the mixture model, selects the number of components and defines the respective methylation states.

# Usage

```
MethylMix_ModelSingleGene(GeneName, METdataVector, METdataNormalVector = NULL,
NoNormalMode = FALSE, maxComp = 3, PvalueThreshold = 0.01,
MeanDifferenceTreshold = 0.1, minSamplesPerGroup = 1)
```

### **Arguments**

GeneName character string with the name of the gene to model METdataVector vector with methylation data for cancer samples.

METdataNormalVector

vector with methylation data for normal samples. It can be NULL and then no

normal mode will be used.

NoNormalMode logical, if TRUE no comparison to normal samples is performed. Defaults to

FALSE.

maxComp maximum number of mixture components admitted in the model (3 by default).

PvalueThreshold

threshold to consider results significant.

MeanDifferenceTreshold

threshold in beta value scale from which two methylation means are considered

different.

minSamplesPerGroup

minimum number of samples required to belong to a new mixture component in order to accept it. Defaul is 1 (not used). If -1, each component has to have at least 5% of all cancer samples.

#### **Details**

maxComp, PvalueThreshold, METDiffThreshold, minSamplesPerGroup are arguments for this function but are fixed in their default values for the user because they are not available in the main MethylMix function, to keep it simple. It would be easy to make them available to the user if we want to.

#### Value

NrComponents number of components identified.

Models an object with the parameters of the model fitted.

MethylationStates vector with DM values for each sample.

MixtureStates vector with DMvalues for each component.

Classifications a vector indicating to which component each sample was assigned.

FlipOverState FlipOverState

MethylMix\_PlotModel The MethylMix\_PlotModel function.

#### **Description**

Produces plots to represent MethylMix's output.

#### Usage

```
MethylMix_PlotModel(GeneName, MixtureModelResults, METcancer, GEcancer = NULL,
    METnormal = NULL)
```

#### **Arguments**

GeneName Name of the gene for which to create a MethylMix plot.

MixtureModelResults

List returned by MethylMix function.

METcancer Matrix with the methylation data of cancer tissue with genes in rows and samples

in columns.

Gene expression data for cancer tissue with genes in rows and samples in columns

(optional).

METnormal Matrix with the normal methylation data of the same genes as in METcancer

(optional). Again genes in rows and samples in columns.

# Value

a list with MethylMix plots, a histogram of the methylation data (MixtureModelPlot) and a scatterplot between DNA methylation and gene expression (CorrelationPlot, is NULL if gene expression data is not provided). Both plots show the different mixture components identified.

# **Examples**

```
# Load the three data sets needed for MethylMix
data(METcancer)
data(METnormal)
data(GEcancer)

# Run methylmix on a small set of example data
MethylMixResults <- MethylMix(METcancer, GEcancer, METnormal)
# Plot the most famous methylated gene for glioblastoma</pre>
```

MethylMix\_Predict 17

```
MethylMix_PlotModel("MGMT", MethylMixResults, METcancer)

# Plot MGMT also with its normal methylation variation
MethylMix_PlotModel("MGMT", MethylMixResults, METcancer, METnormal = METnormal)

# Plot a MethylMix model for another gene
MethylMix_PlotModel("ZNF217", MethylMixResults, METcancer, METnormal = METnormal)

# Also plot the inverse correlation with gene expression (creates two separate plots)
MethylMix_PlotModel("MGMT", MethylMixResults, METcancer, GEcancer, METnormal)

# Plot all functional and differential genes
for (gene in MethylMixResults$MethylationDrivers) {
    MethylMix_PlotModel(gene, MethylMixResults, METcancer, METnormal = METnormal)
}
```

MethylMix\_Predict

The MethylMix\_Predict function

### **Description**

Given a new data set with methylation data, this function predicts the mixture component for each new sample and driver gene. Predictions are based on posterior probabilities calculated with MethylMix'x fitted mixture model.

### Usage

MethylMix\_Predict(newBetaValuesMatrix, MethylMixResult)

#### **Arguments**

newBetaValuesMatrix

Matrix with new observations for prediction, genes/cpg sites in rows, samples in columns. Although this new matrix can have a different number of genes/cpg sites than the one provided as METcancer when running MethylMix, naming of genes/cpg sites should be the same.

MethylMixResult

Output object from MethylMix

## Value

A matrix with predictions (indices of mixture component), driver genes in rows, new samples in columns

# **Examples**

```
# load the three data sets needed for MethylMix
data(METcancer)
data(METnormal)
data(GEcancer)
# run MethylMix on a small set of example data
```

```
MethylMixResults <- MethylMix(METcancer, GEcancer, METnormal)
# toy example new data, of same dimension of original METcancer data
newMETData <- matrix(runif(length(METcancer)), nrow = nrow(METcancer))
rownames(newMETData) <- rownames(METcancer)
colnames(newMETData) <- paste0("sample", 1:ncol(METcancer))
predictions <- MethylMix_Predict(newMETData, MethylMixResults)</pre>
```

MethylMix\_RemoveFlipOver

The MethylMix\_RemoveFlipOver function

### **Description**

Internal. The estimated densities for each beta component can overlap, generating samples that look like being separated from their group. This function re classifies such samples.

### Usage

```
MethylMix_RemoveFlipOver(OrigOrder, MethylationState, classification,
    METdataVector, NrComponents, UseTrainedFlipOver = FALSE,
    FlipOverState = 0)
```

### Arguments

OrigOrder order of sorted values in the methylation vector.

 ${\tt MethylationState}$ 

methylation states for this gene.

classification vector with integers indicating to wich component each sample was classified

into.

METdataVector vector with methylation values from the cancer samples.

NrComponents number of components in this gene.

 ${\tt UseTrainedFlipOver}$ 

FlipOverState

# Value

Corrected vectors with methylation states and classification.

METnormal 19

METnormal DNA methylation data from normal tissue from glioblastoma patients

#### **Description**

Normal tissue DNA methylation data of glioblastoma patients. These normal tissue samples were run on the same platform and are described in the publication referenced below.

### Usage

data(METnormal)

#### **Format**

A numeric matrix with 14 rows (genes) and 4 columns (samples).

#### References

Noushmehr H, Weisenberger DJ, Diefes K, Phillips HS, Pujara K, Berman BP, Pan F, Pelloski CE, Sulman EP, Bhat KP, Verhaak RG, Hoadley KA, Hayes DN, Perou CM, Schmidt HK, Ding L, Wilson RK, Van Den Berg D, Shen H, Bengtsson H, Neuvial P, Cope LM, Buckley J, Herman JG, Baylin SB, Laird PW, Aldape K; Cancer Genome Atlas Research Network. Identification of a CpG island methylator phenotype that defines a distinct subgroup of glioma. Cancer Cell. 2010 May 18;17(5):510-22. doi: 10.1016/j.ccr.2010.03.017. Epub 2010 Apr 15. PubMed PMID: 20399149; PubMed Central PMCID: PMC2872684

predictOneGene

The predictOneGene function

#### **Description**

Auxiliar function. Given a new vector of beta values, this function calculates a matrix with posterior prob of belonging to each mixture commponent (columns) for each new beta value (rows), and return the number of the mixture component with highest posterior probabilit

#### Usage

predictOneGene(newVector, mixtureModel)

### **Arguments**

newVector vector with new beta values

mixtureModel beta mixture model object for the gene being evaluated.

# Value

A matrix with predictions (indices of mixture component), driver genes in rows, new samples in columns

Preprocess\_CancerSite\_Methylation27k

The Preprocess\_CancerSite\_Methylation27k function

# **Description**

Internal. Pre-processes DNA methylation data from TCGA from Illymina 27k arrays.

### Usage

```
Preprocess_CancerSite_Methylation27k(CancerSite, METdirectory,
    MissingValueThreshold = 0.2)
```

#### **Arguments**

CancerSite character of length 1 with TCGA cancer code.

METdirectory character with directory where a folder for downloaded files will be created. Can

be the object returned by the Download\_DNAmethylation function.

MissingValueThreshold

threshold for removing samples or genes with missing values.

#### Value

List with pre processed methylation data for cancer and normal samples.

Preprocess\_CancerSite\_Methylation450k

The Preprocess\_CancerSite\_Methylation450k function

### **Description**

Internal. Pre-processes DNA methylation data from TCGA from Illymina 450k arrays.

# Usage

```
Preprocess_CancerSite_Methylation450k(CancerSite, METdirectory,
    MissingValueThreshold = 0.2)
```

#### **Arguments**

CancerSite character of length 1 with TCGA cancer code.

 ${\tt MET directory} \qquad \text{character with directory where a folder for downloaded files will be created. Can}$ 

be the object returned by the Download\_DNAmethylation function.

 ${\tt MissingValueThreshold}$ 

threshold for removing samples or genes with missing values.

# Value

List with pre processed methylation data for cancer and normal samples.

Preprocess\_DNAmethylation

The Preprocess\_DNAmethylation function

### **Description**

Pre-processes DNA methylation data from TCGA.

# Usage

```
Preprocess_DNAmethylation(CancerSite, METdirectories,
   MissingValueThreshold = 0.2)
```

# Arguments

CancerSite character of length 1 with TCGA cancer code.

METdirectories character vector with directories with the downloaded data. It can be the object returned by the Download\_DNAmethylation function.

MissingValueThreshold

threshold for removing samples or genes with missing values.

#### Details

Pre-process includes eliminating samples and genes with too many NAs, imputing NAs, and doing Batch correction. If there is both 27k and 450k data, and both data sets have more than 50 samples, we combine the data sets, by reducing the 450k data to the probes present in the 27k data, and bath correction is performed again to the combined data set. If there are samples with both 27k and 450k data, the 450k data is used and the 27k data is discarded, before the step mentioned above. If the 27k or the 450k data does not have more than 50 samples, we use the one with the greatest number of samples, we do not combine the data sets.

# Value

List with the pre-processed data matrix for cancer and normal samples.

# **Examples**

```
## Not run:

# Optional register cluster to run in parallel
library(doParallel)
cl <- makeCluster(5)
registerDoParallel(cl)

# Methylation data for ovarian cancer
cancerSite <- "OV"
targetDirectory <- paste0(getwd(), "/")

# Downloading methylation data
METdirectories <- Download_DNAmethylation(cancerSite, targetDirectory, TRUE)
# Processing methylation data</pre>
```

```
METProcessedData <- Preprocess_DNAmethylation(cancerSite, METdirectories)

# Saving methylation processed data
saveRDS(METProcessedData, file = paste0(targetDirectory, "MET_", cancerSite, "_Processed.rds"))

# Clustering methylation data
res <- ClusterProbes(METProcessedData[[1]], METProcessedData[[2]])

# Saving methylation clustered data
toSave <- list(METcancer = res[[1]], METnormal = res[[2]], ProbeMapping = res$ProbeMapping)
saveRDS(toSave, file = paste0(targetDirectory, "MET_", cancerSite, "_Clustered.rds"))

stopCluster(cl)

## End(Not run)</pre>
```

Preprocess\_GeneExpression

The Preprocess\_GeneExpression function

### **Description**

Pre-processes gene expression data from TCGA.

### Usage

```
Preprocess_GeneExpression(CancerSite, MAdirectories,
   MissingValueThresholdGene = 0.3, MissingValueThresholdSample = 0.1)
```

# Arguments

CancerSite character of length 1 with TCGA cancer code.

MAdirectories character vector with directories with the downloaded data. It can be the object

returned by the Download\_DNAmethylation function.

 ${\tt MissingValueThresholdGene}$ 

threshold for missing values per gene. Genes with a percentage of NAs greater

than this threshold are removed. Default is 0.3.

MissingValueThresholdSample

threshold for missing values per sample. Samples with a percentage of NAs

greater than this threshold are removed. Default is 0.1.

#### **Details**

Pre-process includes eliminating samples and genes with too many NAs, imputing NAs, and doing Batch correction.

# Value

List with the pre-processed data matrix for cancer and normal samples.

#### **Examples**

```
## Not run:
# Optional register cluster to run in parallel
library(doParallel)
cl <- makeCluster(5)</pre>
registerDoParallel(cl)
# Gene expression data for ovarian cancer
cancerSite <- "OV"
targetDirectory <- paste0(getwd(), "/")</pre>
# Downloading gene expression data
GEdirectories <- Download_GeneExpression(cancerSite, targetDirectory, TRUE)
# Processing gene expression data
GEProcessedData <- Preprocess_GeneExpression(cancerSite, GEdirectories)</pre>
# Saving gene expression processed data
saveRDS(GEProcessedData, file = paste0(targetDirectory, "GE_", cancerSite, "_Processed.rds"))
stopCluster(cl)
## End(Not run)
```

Preprocess\_MAdata\_Cancer

 $The \ Preprocess\_MA data\_Cancer \ function$ 

# **Description**

Internal. Pre-process gene expression data for cancer samples.

### Usage

```
Preprocess_MAdata_Cancer(CancerSite, Directory, File,
   MissingValueThresholdGene = 0.3, MissingValueThresholdSample = 0.1)
```

# **Arguments**

CancerSite TCGA code for the cancer site.

Directory Directory.
File File.
MissingValueThresholdGene

threshold for missing values per gene. Genes with a percentage of NAs greater

than this threshold are removed. Default is 0.3.

MissingValueThresholdSample

threshold for missing values per sample. Samples with a percentage of NAs greater than this threshold are removed. Default is 0.1.

Value

The data matrix.

24 SNPprobes

Preprocess\_MAdata\_Normal

The Preprocess\_MAdata\_Normal function

# Description

Internal. Pre-process gene expression data for normal samples.

# Usage

```
Preprocess_MAdata_Normal(CancerSite, Directory, File,
   MissingValueThresholdGene = 0.3, MissingValueThresholdSample = 0.1)
```

## **Arguments**

CancerSite TCGA code for the cancer site.

Directory Directory.
File File.

 ${\tt MissingValueThresholdGene}$ 

threshold for missing values per gene. Genes with a percentage of NAs greater

than this threshold are removed. Default is 0.3.

 ${\tt Missing Value Threshold Sample}$ 

threshold for missing values per sample. Samples with a percentage of NAs

greater than this threshold are removed. Default is 0.1.

## Value

The data matrix.

ProbeAnnotation ProbeAnnotation data set

# Description

Data set with annotation from Illumina methylatin arrays mapping CpG sites to genes.

SNPprobes SNPprobes data set

### **Description**

Vector with probes with SNPs.

 ${\tt TCGA\_BatchCorrection\_MolecularData}$ 

 $The\ TCGA\_Batch Correction\_Molecular Data\ function$ 

# Description

Internal. Wrapper to perform batch correction.

# Usage

TCGA\_BatchCorrection\_MolecularData(GEN\_Data, BatchData, MinInBatch)

# Arguments

GEN\_Data matrix with data to be corrected for batch effects.

BatchData Batch data.

MinInBatch minimum number of samples per batch.

# Value

The corrected data matrix.

TCGA\_GENERIC\_BatchCorrection

The TCGA\_GENERIC\_BatchCorrection function

# Description

Internal. Performs batch correction.

# Usage

TCGA\_GENERIC\_BatchCorrection(GEN\_Data, BatchData)

# Arguments

GEN\_Data matrix with data to be corrected for batch effects.

BatchData Batch data.

### Value

The corrected data matrix.

TCGA\_GENERIC\_CheckBatchEffect

The TCGA\_GENERIC\_CheckBatchEffect function

# Description

Internal. Checks if batch correction is needed.

# Usage

TCGA\_GENERIC\_CheckBatchEffect(GEN\_Data, BatchData)

# Arguments

GEN\_Data matrix with data to be corrected for batch effects.

BatchData Batch data.

#### Value

list with results.

 ${\tt TCGA\_GENERIC\_CleanUpSampleNames}$ 

 $The\ TCGA\_GENERIC\_CleanUpSampleNames\ function$ 

# Description

Internal. Cleans the samples IDs into the 12 digit format and removes doubles.

# Usage

TCGA\_GENERIC\_CleanUpSampleNames(GEN\_Data, IDlength = 12)

# Arguments

GEN\_Data data matrix.

IDlength length of samples ID.

### Value

data matrix with cleaned sample names.

TCGA\_GENERIC\_GetSampleGroups

The TCGA\_GENERIC\_GetSampleGroups function

# **Description**

Internal. Looks for the group of the samples (normal/cancer).

# Usage

TCGA\_GENERIC\_GetSampleGroups(SampleNames)

# **Arguments**

SampleNames vector with sample names.

#### Value

a list.

 ${\tt TCGA\_GENERIC\_LoadIlluminaMethylationData}$ 

The TCGA\_GENERIC\_LoadIlluminaMethylationData function

# Description

Internal. Read in an illumina methylation file with the following format: header row with sample labels, 2nd header row with 4 columns per sample: beta-value, geneSymbol, chromosome and GenomicCoordinate. The first column has the probe names.

# Usage

TCGA\_GENERIC\_LoadIlluminaMethylationData(Filename)

# Arguments

Filename

name of the file with the data.

# Value

methylation data.

TCGA\_GENERIC\_MergeData

The TCGA\_GENERIC\_MergeData function

### **Description**

Internal.

### Usage

TCGA\_GENERIC\_MergeData(NewIDListUnique, DataMatrix)

#### **Arguments**

NewIDListUnique

unique rownames of data.

DataMatrix data matrix.

#### Value

data matrix.

 $\label{thm:constraint} TCGA\_GENERIC\_MET\_ClusterProbes\_Helper\_ClusterGenes\_with\_hclust\\ The\ TCGA\_GENERIC\_MET\_ClusterProbes\_Helper\_ClusterGenes\_with\_hclust\\ function$ 

# Description

Internal. Cluster probes into genes.

### Usage

```
TCGA_GENERIC_MET_ClusterProbes_Helper_ClusterGenes_with_hclust(Gene,
    ProbeAnnotation, MET_Cancer, MET_Normal = NULL, CorThreshold = 0.4)
```

# **Arguments**

Gene gene.

 ${\tt ProbeAnnotation}$ 

data set matching probes to genes.

MET\_Cancer data matrix for cancer samples.

MET\_Normal data matrix for normal samples.

CorThreshold correlation threshold for cutting the clusters.

### Value

List with the clustered data sets and the mapping between probes and genes.

TCGA\_Load\_MolecularData

The TCGA\_Load\_MolecularData function

# **Description**

Internal. Reads in gene expressiondata. Deletes samples and genes with more NAs than the respective thresholds. Imputes other NAs values.

# Usage

```
TCGA_Load_MolecularData(Filename, MissingValueThresholdGene = 0.3,
    MissingValueThresholdSample = 0.1)
```

#### **Arguments**

Filename name of the file with the data.

MissingValueThresholdGene

threshold for missing values per gene. Genes with a percentage of NAs greater than this threshold are removed. Default is 0.3.

MissingValueThresholdSample

threshold for missing values per sample. Samples with a percentage of NAs greater than this threshold are removed. Default is 0.1.

#### Value

gene expression data.

TCGA\_Process\_EstimateMissingValues

The TCGA\_Process\_EstimateMissingValues function

# Description

Internal. Removes patients and genes with more missing values than the Missing Value Threshold, and imputes remaining missing values using Tibshirani's KNN method.

# Usage

TCGA\_Process\_EstimateMissingValues(MET\_Data, MissingValueThreshold = 0.2)

# **Arguments**

MET\_Data data matrix. MissingValueThreshold

threshold for removing samples and genes with too many missing values.

#### Value

the data set with imputed values and possibly some genes or samples deleted.

# **Index**

```
* cluster
                                                                                                                    TCGA_Process_EstimateMissingValues,
         GetData, 9
                                                                                                          * preprocess
* cluter_probes
                                                                                                                    GetData, 9
         ClusterProbes, 4
                                                                                                                    Preprocess_DNAmethylation, 21
* datasets
                                                                                                                    Preprocess_GeneExpression, 22
         BatchData, 3
         GEcancer, 8
                                                                                                          BatchData, 3
         METcancer, 11
                                                                                                          betaEst_2, 3
         METnormal, 19
                                                                                                          blc_2, 3
         ProbeAnnotation, 24
         SNPprobes, 24
                                                                                                          ClusterProbes, 4
* download
                                                                                                          ComBat_NoFiles, 5
         Download_DNAmethylation, 6
                                                                                                          combineForEachOutput, 6
         Download_GeneExpression, 7
         GetData, 9
                                                                                                          Download_DNAmethylation, 6
* internal
                                                                                                          Download_GeneExpression, 7
         betaEst_2, 3
         blc_2, 3
                                                                                                          GEcancer. 8
         ComBat_NoFiles, 5
                                                                                                          get_firehoseData, 10
         combineForEachOutput, 6
                                                                                                          GetData, 9
         get_firehoseData, 10
         MethylMix_MixtureModel, 13
                                                                                                          METcancer, 11
         MethylMix_ModelSingleGene, 15
                                                                                                          MethylMix, 12
         MethylMix_RemoveFlipOver, 18
                                                                                                          MethylMix_MixtureModel, 13
         Preprocess_CancerSite_Methylation27k,
                                                                                                          MethylMix_ModelGeneExpression, 14
                                                                                                          MethylMix_ModelSingleGene, 15
         Preprocess_CancerSite_Methylation450k,
                                                                                                          MethylMix_PlotModel, 16
                                                                                                          MethylMix_Predict, 17
         Preprocess_MAdata_Cancer, 23
                                                                                                          MethylMix_RemoveFlipOver, 18
         {\tt Preprocess\_MAdata\_Normal}, 24
                                                                                                          METnormal, 19
         TCGA_BatchCorrection_MolecularData,
                                                                                                          predictOneGene, 19
         TCGA_GENERIC_BatchCorrection, 25
                                                                                                          Preprocess_CancerSite_Methylation27k,
         TCGA_GENERIC_CheckBatchEffect, 26
         TCGA_GENERIC_CleanUpSampleNames,
                                                                                                          Preprocess_CancerSite_Methylation450k,
         TCGA_GENERIC_GetSampleGroups, 27
                                                                                                          Preprocess_DNAmethylation, 21
         {\tt TCGA\_GENERIC\_LoadIlluminaMethylationData, Preprocess\_Gene Expression, 22}
                  27
                                                                                                          Preprocess_MAdata_Cancer, 23
         TCGA_GENERIC_MergeData, 28
                                                                                                          Preprocess_MAdata_Normal, 24
         TCGA_GENERIC_MET_ClusterProbes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes_Helper_ClusterDes
         TCGA_Load_MolecularData, 29
                                                                                                          SNPprobes, 24
```

INDEX 31

```
TCGA_BatchCorrection_MolecularData, 25
TCGA_GENERIC_BatchCorrection, 25
TCGA_GENERIC_CheckBatchEffect, 26
TCGA_GENERIC_CleanUpSampleNames, 26
TCGA_GENERIC_GetSampleGroups, 27
TCGA_GENERIC_LoadIlluminaMethylationData, 27
TCGA_GENERIC_MergeData, 28
TCGA_GENERIC_MET_ClusterProbes_Helper_ClusterGenes_with_hclust, 28
TCGA_Load_MolecularData, 29
TCGA_Process_EstimateMissingValues, 29
```