# Bioconductor for highthroughput genetic data

分析处理高通量基因数据: Bioconductor

Martin Morgan martin.morgan@roswellpark.org 29 May 2016



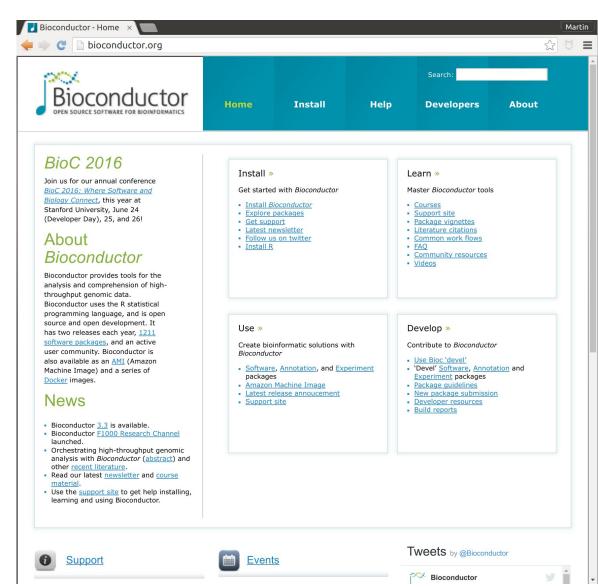


### Bioconductor

Statistical analysis and comprehension of high-throughput genomic data 统计学方法分析理解高通量基因 组数据

- Sequence (RNA-seq, ChIP-seq, Variants, ...)
- Microarrays (expression, methylation, SNP, copy number, ...)
- Flow cytometry
- Proteomics
- Image analysis
- ...

https://bioconductor.org/ https://support.bioconductor.org



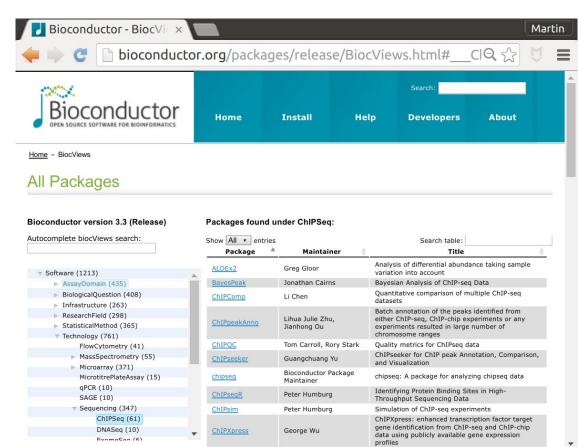
### **Bioconductor**

1211 Software Packages 1211个软件包

- All packages
   https://bioconductor.org/packages
- Example package: <u>GenomicRanges</u>

#### Features 特征

- Vignettes and support site for learning
- Stable 'release' branch for users;
   reproducible research
- 'Devel' branch for new features & packages
- Classes for inter-operability between packages



source("https://bioconductor.org/biocLite.R")
biocLite("GenomicRanges")

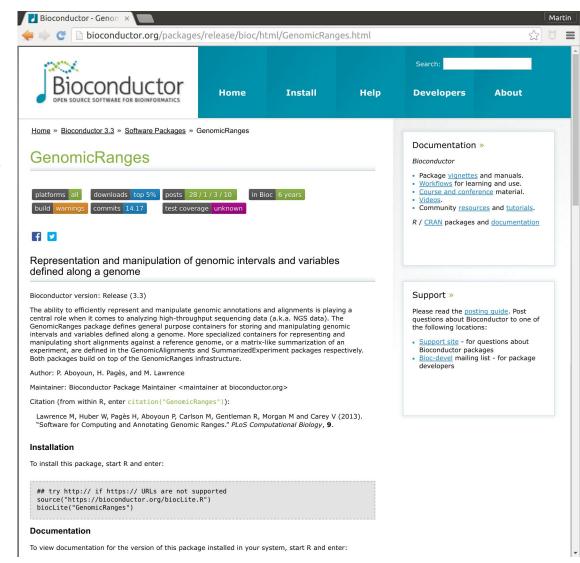
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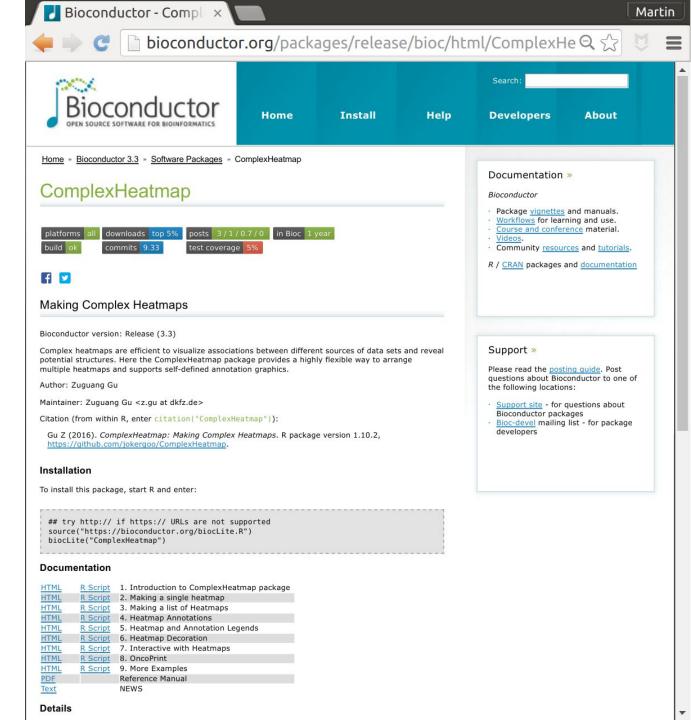
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Vignettes 文档说明

Each package!

Courses 相关课程

Support site



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Support site 支持网站

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1		ndard workflow		4			
	1.1						
1.2 How to get help							
	1.3						
		,					
1.3.3 Count matrix input							
		Description of the second seco	to gene-level				
		1.3.5 <i>HTSeq</i> input					
		1.3.7 Note on factor level	S	q	1		
		1.3.8 Collapsing technica	Differential a				
		1.3.9 About the pasilla d	Dilicicitiai a	nalysis of count data – the DESeq2 package			
	1.4	Differential expression analy					
1.5 Exploring and exporting res			Mich	ael I. Love <sup>1</sup> , Simon Anders <sup>2,3</sup> , Wolfgang Huber <sup>3</sup>			
		1.5.1 MA-plot					
1.5.2 Plot counts 1 Depa				rtment of Biostatistics, Dana-Farber Cancer Institute and			
	1.5.3 More information o			rvard TH Chan School of Public Health, Boston, US;			
1.5.4 Rich visualization a <sup>2</sup> Institut				e for Molecular Medicine Finland (FIMM), Helsinki, Finland;			
		1.5.5 Exporting results to	<sup>3</sup> European	Molecular Biology Laboratory (EMBL), Heidelberg, Germany			
	1.6	Multi-factor designs					
				May 15, 2016			
2	2 Data transformations and visu						
2.1 Count data transformations Abst				Abstract			
		2.1.1 Blind dispersion est	A basic task in the ana	lysis of count data from RNA-seq is the detection of differentially expressed			
		2.1.2 Extracting transform		presented as a table which reports, for each sample, the number of sequence			
		2.1.3 Regularized log trai		signed to each gene. Analogous data also arise for other assay types, including			
		2.1.4 Variance stabilizing		, shRNA screening, mass spectrometry. An important analysis question is the			
		2.1.5 Effects of transform		I inference of systematic changes between conditions, as compared to within- backage DESeq2 provides methods to test for differential expression by use			
	2.2	Data quality assessment by		lized linear models; the estimates of dispersion and logarithmic fold changes			
		2.2.1 Heatmap of the cou		distributions <sup>1</sup> . This vignette explains the use of the package and demonstrates			
		2.2.2 Heatmap of the sar	typical workflows. An RN	Differential analysis of count data – the DESeq2 package	4		
		2.2.3 Principal componer	vignette but at a slower pac	Differential analysis of count data – the DESeq2 package	4		
			DESeq2 version: 1.12.				
3	Vari	ations to the standard wo	KITOW	1 Standard workflow			
	3.1 Wald test individual steps						
	3.2	Contrasts		1.1 Quick start			
	3.3 Interactions				and the second second second second second second		
	3.4 Time-series experiments			Here we show the most basic steps for a differential expression analysis. These steps require you have a RangedSummarizedExperiment object se which contains the counts and information about samples. The design indicates that we want to measure the effect of condition, controlling for batch differences. The two			
3.5 Likelihood ratio test							
				factor variables batch and condition should be columns of colData(se).			
				dds <- DESeqDataSet(se, design = ~ batch + condition)			
				dds <- DESeq(dds)			

res <- results(dds, contrast=c("condition","trt","con"))</pre>

instead of DESeqDataSet, as shown in Section 1.3.3.

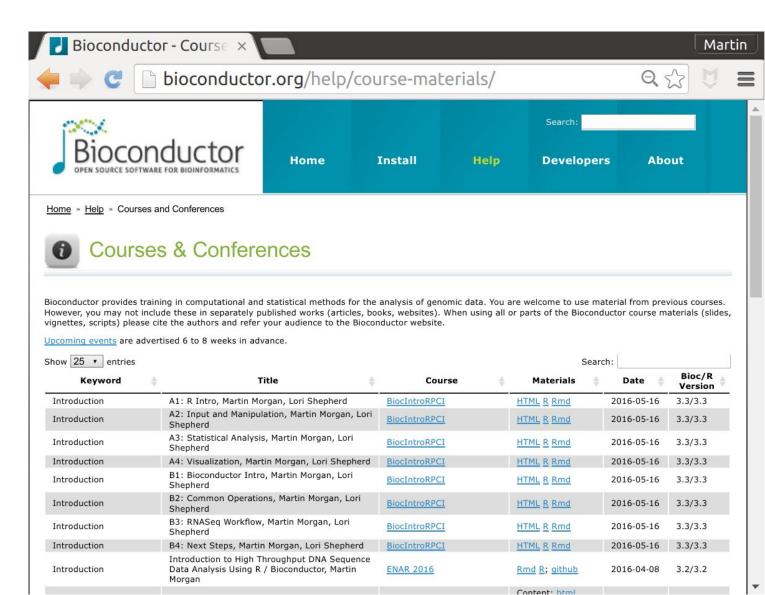
If you have a count matrix and sample information table, the first line would use DESeqDataSetFromMatrix

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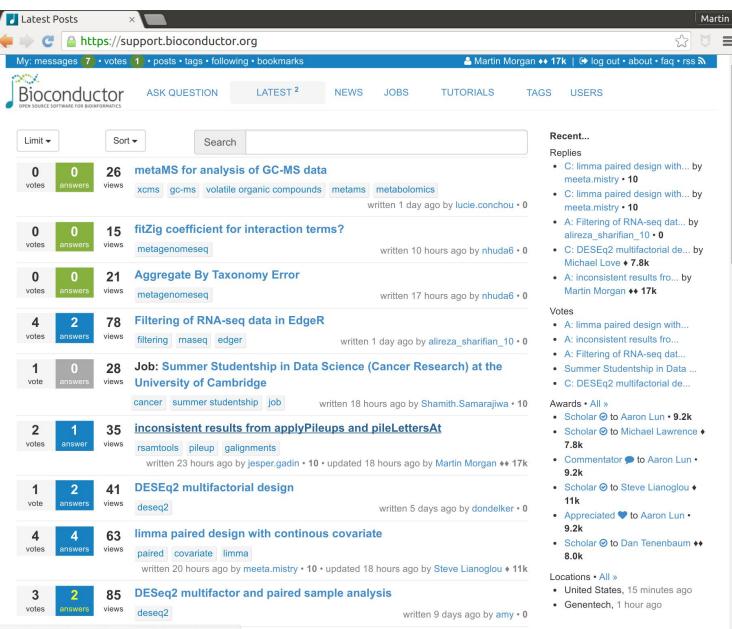


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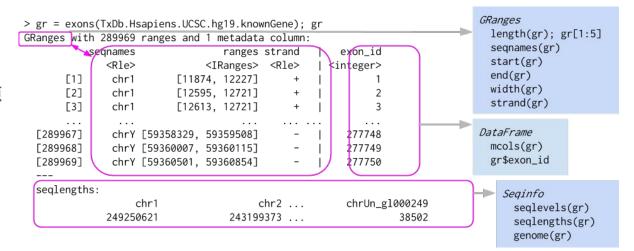
https://support.bioconductor.org/p/83141/differential binding above a log2 fold change threshold using

#### Essential Software 基础软件包

- Biostrings
- GenomicRanges
- SummarizedExperiment

#### Annotation Resources 注释资源

- Packages
  - o org.\*
  - o TxDb.\*
  - o BSgenome.\*
- On-line
  - o biomaRt
  - AnnotationHub

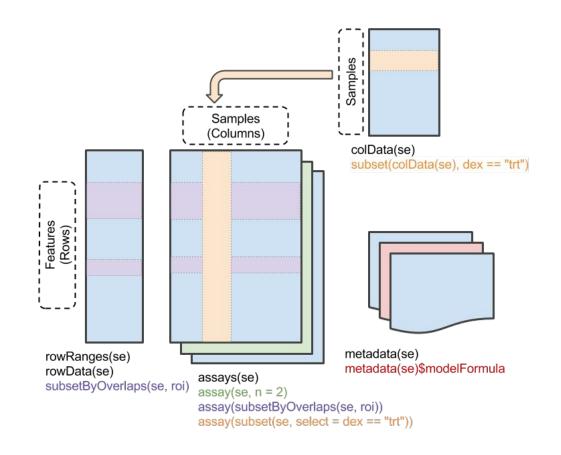


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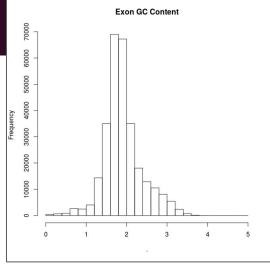
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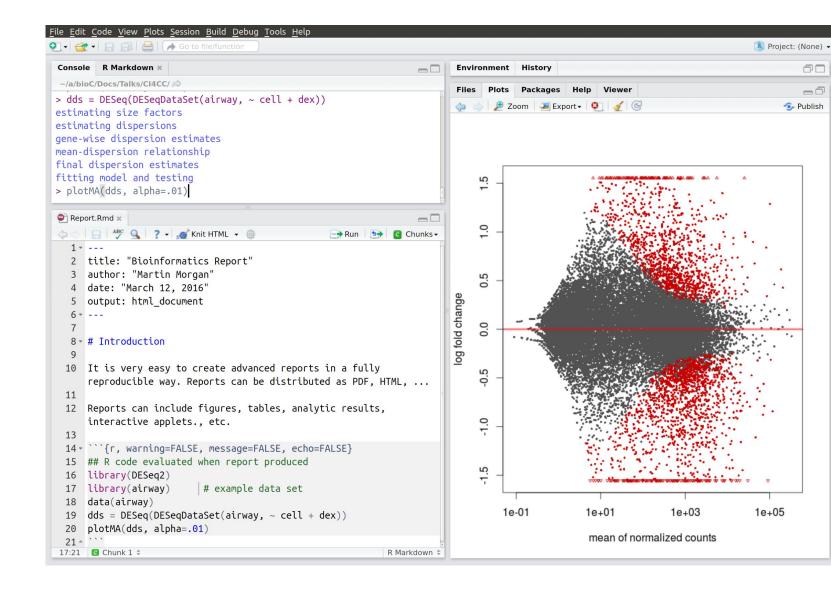
#### Annotation Resources 注释资源

- Packages
  - Identifier mapping
  - Gene models
  - Genome sequence
- On-line
  - biomaRt
  - AnnotationHub

```
library(Homo.sapiens)
 head(id <- select(Homo.sapiens, "BRCA1", c("GENENAME", "TXID"), "SYMBOL"), 3)
 select()' returned 1:many mapping between keys and columns
                GENENAME TXID
 SYMBOL
  BRCA1 breast cancer 1 63594
  BRCA1 breast cancer 1 63595
  BRCA1 breast cancer 1 63596
 brca1tx <- exonsBy(Homo.sapiens, "tx")[id$TXID]</pre>
  library(BSgenome.Hsapiens.UCSC.hg19)
 head(dna <- getSeq(BSgenome.Hsapiens.UCSC.hg19, brca1tx))
DNAStringSetList of length 6
 . 63594"]] TTCATTGGAACAGAAAGAAATGGATTTATCTGCTCTTCGCGTTGAAGAAGTACAAAATGTCATTA
  '63595"]] AGATAACTGGGCCCCTGCGCTCAGGAGGCCTTCACCCTCTGCTCTGGGTAAAGGTAGTAGAGTCC.
  "63596"]] AGATAACTGGGCCCCTGCGCTCAGGAGGCCTTCACCCTCTGCTCTGGGTAAAGGTAGTAGAGTCC
  "63597"]] CTTAGCGGTAGCCCCTTGGTTTCCGTGGCAACGGAAAAGCGCGGGAATTACAGATAAATTAAAAC
  "63598"]] CTTAGCGGTAGCCCCTTGGTTTCCGTGGCAACGGAAAAGCGCGGGAATTACAGATAAATTAAAAC..
  "63599"]] GTACCTTGATTTCGTATTCTGAGAGGCTGCTGCTTAGCGGTAGCCCCTTGGTTTCCGTGGCAACG...
 library(magrittr)
  getSeq(BSgenome.Hsapiens.UCSC.hg19, exons(Homo.sapiens)) %>%
      letterFrequency("GC") %>% log10 %>% hist(main="Exon GC Content")
                                               Exon GC Content
```



### Use



### Use

### Gene expression 基因表达

- RNA-seq: DESeq2, edgeR, scde, ...
- Microarray: limma

### Gene regulation 基因调控

- ChIP-seq: csaw, DiffBind
- Methylation arrays: minfi, missMethyl
- Gene set enrichment: topGO, limma

#### Variants 变异

- VariantAnnotation
- VariantFiltering

### Flow cytometry 流式细胞仪

flowCore

#### Data access 数据访问

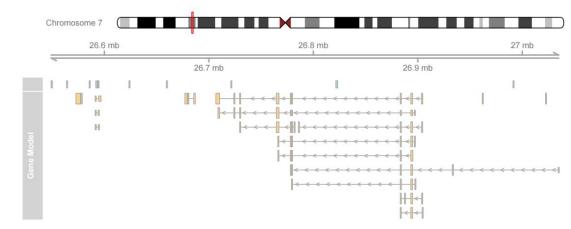
- biomaRt
- GEOquery / SRAdb
- TCGAbiolinks
- AnnotationHub / ExperimentHub

#### Visualization 可视化

• Gviz, ComplexHeatmap, ggbio, ggtree, ...

### Many other packages!

> plotTracks(list(itrack, gtrack, atrack, grtrack))



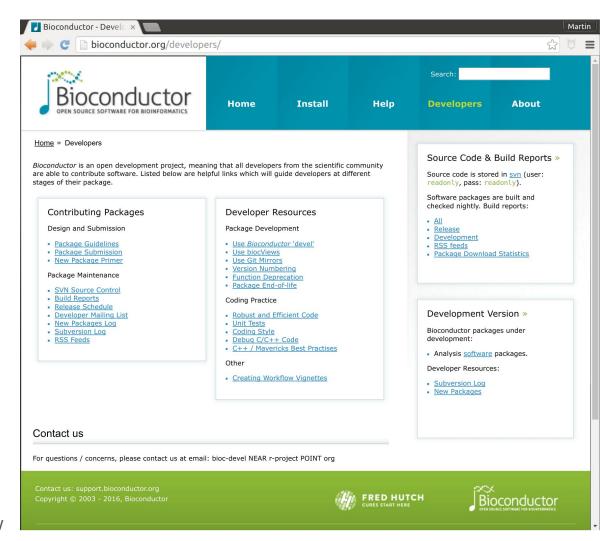
## Develop

### Create a package 创建包

- Use existing classes --DNAStringSet, GRanges, SummarizedExperiment
- Use existing packages -rtracklayer, Rsamtools, Biostrings, ...
- Full vignette and examples
- <u>Unit tests</u> and other best practices

#### Submit to Bioconductor 提交

- Technical review
- Long-term <u>support</u> & maintenance
- Introduced to 'devel' branch, new release every April and October



# Acknowledgements

### Core team (current & recent)

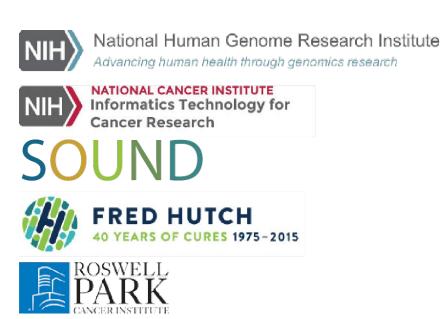
 Valerie Obenchain, Herve Pages, Dan Tenenbaum, Lori Shepherd, Marcel Ramos, Brian Long, Jim Hester, Jim Java, Sonali Arora, Nate Hayden, Paul Shannon, Marc Carlson

### Technical advisory board

 Vincent Carey, Wolfgang Huber, Robert Gentleman, Rafael Irizzary, Levi Waldron, Michael Lawrence, Sean Davis, Aedin Culhane

### Scientific advisory board

 Simon Tavare (CRUK), Paul Flicek (EMBL/EBI), Simon Urbanek (AT&T), Vincent Carey (Brigham & Women's), Wolfgang Huber (EBI), Raphael Irizzary, Robert Gentleman (23andMe)



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### Resources

### Web sites

- https://bioconductor.org
- https://support.bioconductor.org

### **Events**

- Annual Conference, Stanford, CA, USA, 24-26 June
- Bioconductor Asia-Pacific Conference, Brisbane, Australia 3-4 November
- Bioconductor European Developer Workshop, Basel, Switzerland, 6-7
   December