Package 'simpleSeg'

July 3, 2025

Type Package

Title A package to perform simple cell segmentation

Description Image segmentation is the process of identifying the borders of individual objects (in this case cells) within an image. This allows for the features of cells such as marker expression and morphology to be extracted, stored and analysed. simpleSeg provides functionality for user friendly, watershed based segmentation on multiplexed cellular images in R based on the intensity of user specified protein marker channels. simpleSeg can also be used for the normalization of single cell data obtained from multiple images.

```
Version 1.10.1
Date 2022-04-21
VignetteBuilder knitr
Encoding UTF-8
biocViews Classification, Survival, SingleCell, Normalization, Spatial
Imports BiocParallel, EBImage, terra, stats, spatstat.geom, S4Vectors,
     grDevices, SummarizedExperiment, methods, cytomapper
Suggests BiocStyle, testthat (>= 3.0.0), knitr, ggplot2
License GPL-3
RoxygenNote 7.2.3
Config/testthat/edition 3
BugReports https://github.com/SydneyBioX/simpleSeg/issues
URL https://sydneybiox.github.io/simpleSeg/
     https://github.com/SydneyBioX/simpleSeg
git_url https://git.bioconductor.org/packages/simpleSeg
git branch RELEASE 3 21
git last commit b7dc106
git_last_commit_date 2025-06-18
Repository Bioconductor 3.21
Date/Publication 2025-07-02
```

2 normalizeCells

Author Nicolas Canete [aut], Alexander Nicholls [aut], Ellis Patrick [aut, cre]

Maintainer Ellis Patrick <ellis.patrick@sydney.edu.au>

Contents

generateBPParam normalizeCells . simpleSeg																						 					2
generateBPParam	U	Itili	ity	fu	nci	tio	n t	0	ge	ne	ra	te	В	PP	A	RN	1 0	obj	iec	t.	_	_	_		_		_

Description

Utility function to generate BPPARM object.

Usage

```
generateBPParam(cores = 1)
```

Arguments

cores

Desired number of cores for BPPARAM object.

Value

A BPPPARAM object.

normalizeCells	Normalizes and transforms cell data in preparation for clustering (ac-
	$cepts\ data frame,\ Single Cell Experiment\ and\ Spatial Experiment).$

Description

Normalizes and transforms cell data in preparation for clustering (accepts dataframe, SingleCellExperiment and SpatialExperiment).

normalizeCells 3

Usage

```
normalizeCells(
  cells,
  markers = NULL,
  assayIn = NULL,
  assayOut = "norm",
  imageID = "imageID",
  transformation = NULL,
  method = NULL,
  cores = 1
)
```

Arguments

cells	A Dataframe of SingleCellExperiment or SpatialExperiment containing cells and features to be normalized/transformed
markers	A list containing the names of cell markers which will be normalized and/or transformed.
assayIn	If input is a SingleCellExperiment or SpatialExperiment with multiple assays, specify the assay to be normalized and/or transformed.
assay0ut	If input is a SingleCellExperiment or SpatialExperiment, the new of the normalized data.
imageID	If input is a SingleCellExperiment or SpatialExperiment, this is the name of the image ID variable in order to stratify. cells correctly
transformation	The transformation/s to be performed, default is NULL, accepted values: 'asinh' and 'sqrt'.
method	The normalization method/s to be performed, default is NULL, accepted values: 'mean', 'minMax', 'trim99', 'PC1'.
cores	The number or cores for parallel processing.

Value

returns a dataframe with individual cells as rows and features as columns.

Examples

```
library(cytomapper)
data("pancreasSCE")
cells.normalized <- normalizeCells(
  cells = pancreasSCE,
  markers = c("CD99", "PIN", "CD8a", "CDH"),
  assayIn = "counts",
  assayOut = "normCounts",
  imageID = "ImageNb",
  transformation = "asinh",
  method = "trim99"
)</pre>
```

4 simpleSeg

simpleSeg

Perform simple segmentation of multiplexed cellular images

Description

Perform simple segmentation of multiplexed cellular images

Usage

```
simpleSeg(
   image,
   nucleus,
   cellBody = "dilate",
   sizeSelection = 10,
   smooth = 1,
   transform = NULL,
   watershed = "intensity",
   tolerance = NULL,
   ext = 1,
   discSize = 3,
   tissue = NULL,
   pca = FALSE,
   cores = 1
)
```

Arguments

ımage			read into the function.

nucleus The marker or list of markers corresponding to the nuclei.

cellBody Method of cytoplasm identification. Can be 'none', dilate', 'discModel' or the

name of a dedicated cytoplasm marker

sizeSelection Minimum pixels for an object to be recognized as a cell and not noise.

smooth The amount of Gaussian smoothing to be applied to the image/s

transform A transformation or list of transformations and normalizations to be performed

prior to nuclei or cytoplasm identification. Accepted vales: "sqrt", "asinh", "norm99", "maxThresh" and "tissueMask". Tissue mask may be used when the sample does not take up the entirety of the image (typically a circular sample inside the image. When tissue mask is specified the background noise present

outside the sample area is removed).

watershed Method used to perform watersheding. Accepted values: "intensity", "distance"

or "combine".

tolerance The minimum height of the object in the units of image intensity between its

highest point (seed) and the point where it contacts another object (checked for every contact pixel). If the height is smaller than the tolerance, the object will be combined with one of its neighbors, which is the highest. Tolerance should simpleSeg 5

	be chosen according to the range of x. Default value is 1, which is a reasonable value if x comes from distmap.
ext	Radius of the neighborhood in pixels for the detection of neighboring objects. Higher value smooths out small objects.
discSize	The size of dilation around nuclei to create cell disc or capture cytoplasm
tissue	Channels to be used to create the tissue mask if specified in transforms.
pca	Whether to run PCA on aggregated nucleus markers in order to detect the cellular nucclei.
cores	The number or cores for parallel processing or a BPPARAM object

Value

A list of image masks

Examples

```
library(cytomapper)
data("pancreasImages")
masks <- simpleSeg(pancreasImages,
   nucleus = "H3",
   cellBody = "discModel",
   sizeSelection = 8,
   smooth = 1.2,
   transform = "sqrt",
   watershed = "combine",
   tolerance = 1, ext = 1,
   discSize = 3,
   cores = 5
)</pre>
```

Index

```
generateBPParam, 2
normalizeCells, 2
simpleSeg, 4
```